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Induction of Apoptosis with *Cyrtopodion Scabrum* Extract in Colon Cancer Cells: A Preliminary Study on Targeting P53 Signaling Pathway

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Abstract

Background: *Cyrtopodion scabrum* is a kind of lizard, widely distributed in southwestern, central, and eastern Iran. In our previous studies, we showed the selective anticancer properties of *Cyrtopodion scabrum* extract (CSE) against certain gastrointestinal cancer cell lines with no significant harmful effects on the normal cells.

Method: This in vitro study was conducted to investigate the mechanism of action of CSE in SW742 (colon cancer cell line) using western blotting, real-time polymerase chain reaction (PCR), and PI staining/flow cytometry analysis.

Results: Western blot analysis showed that CSE upregulates P53 protein expression significantly in SW742 cancer cell line, while QRT-PCR results revealed that P53 mRNA did not increase accordingly. It is proposed that TP53 protein play its role as a tumor suppressor through the protein stability mechanism and not through the increase in the gene expression. Considering this fact, for TP53 stabilization, two of its target genes, p21 and mdm2, were regulated. We evaluated the mRNA expression of these two genes as well. The obtained data showed a significant increase in both genes by CSE (1000 μ g/ml), suggesting that the induction of G2 cell cycle arrest, which we previously reported, may happen through P53 and P210ver-expression. No significant decrease was observed in apoptosis when SW742 cells were co-treated with CSE and the inhibitor of P53 transcriptional activity, (PFT)- α , indicating that the induction of apoptosis with CSE did not occur through P53-dependent transcriptional activity.

Conclusion: The obtained results herein revealed that the observed anticancer effect of CSE may occur through TP53 upregulation, yet with P53-independent transcriptional activity.

Keywords: Natural anticancer product, Gecko (Gekko) extract, *Cyrtopodion scabrum*, Western blot analysis, Real-time PCR, flow cytometry, P53 signaling pathway

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Introduction

Following heart disease, cancer is one of the main causes of death worldwide¹ and the prevalence of colorectal cancer (CRC) is ranked the third among cancers. CRC is the cause of 9.7% of all deaths in the world.² It has the third and fifth prevalence rank among Iranian women and men, respectively.3 Today, certain therapeutic methods are being utilized to treat cancer patients, including chemotherapy as one of the main powerful treatments. Chemotherapy drugs not only kill the cancer cells, but also intensively affect normal cells; therefore, these types of drugs have several unwanted side-effects. To date, many scientists have proposed the use of less toxic and safer drugs, such as natural agents. The use of natural agents, for instance, herbal extracts or animal products and bodies, is one of the traditional medicine (TM) branches. In traditional Chinese medicine (TCM), the whole-body extract of a kind of Chinese gecko (Swinhonis Guenther) has been applied as a drug to treat various diseases, like cancer, for hundreds of years. In our previous studies, we also showed that a kind of gecko, named Cyrtopodion scabrum (C. scabrum) (socalled rough-tailed home gecko), has anti-proliferative effects on the breast, colorectal,⁷ stomach, and liver cancer cells.⁸ C. scabrum is distributed around the Indus Valley to the eastern borders of the Caspian Sea, including Iran, Turkey, Pakistan, Iraq, and so forth.9 The current study aimed to investigate the potential effects of Cyrtopodion scabrum extract (CSE) on the P53 protein level and gene expression, as well as the role of P53 transcriptional pathway in CSE apoptosis induction. The gene expression of p21 and mdm2, as the two target genes of P53 protein pathway, in response to CSE treatment was also investigated.

Materials and Methods

In the present experimental in-vitro research, we studied the effect of CSE on apoptosis induction and the involvement of some elements of P53 protein pathway.

Preparation of aqueous extract from animals

C. scabrum was provided by Razi Research Institute of Vaccine and Serum, Shiraz, Iran. The extract (CSE) was prepared exactly according to the protocol described previously by Amiri et al.⁷ *Cell culture*

SW742 colon cancer cell line was purchased from the National Cell Bank of Pasteur Institute of Iran. The cells were cultured in RPMI-1640 (shellmax, China) supplemented with 10% fetal bovine serum (FBS), 100 U/ml of Penicillin, 100mg/ml of Streptomycin, and were incubated at 37°C in a 5% CO2 incubator.

Western blotting

The cells were treated for 72 hours with complete medium for the control and complete medium containing three different concentrations of CSE (IC50: 250, 2×IC50:500 and 4×IC50: 1000µg/ml) for the tests. According to our previous study, IC50 for SW742 cells was calculated as 251±13µg/ml.⁸ The cell lysates were then prepared and the total protein concentrations were determined using Bradford assay. The samples were separated on a 12% SDS-PAGE gel and then transferred to the nitrocellulose membranes for western blotting. The following antibodies were utilized for probing: TP53 Antibody (Biolegend BTP53-12, UK) for tests and β -actin (Abcam ab6276, USA) for controls. Image J software analysis was employed to determine the band intensity.¹⁰

Quantitative real-time polymerase chain reaction

 2.6×105 SW742 cells were seeded into 25 cm² (T-25) cell culture flasks and after 12 hours of incubation, the cells were treated with different concentrations of CSE for 72h, as mentioned in section 2.3. Following the total RNA extraction using the corresponding mini kit (Yekta Tajhiz Azma, Iran), cDNAs were synthesized with cDNA synthesis kit (Thermo Scientific, USA). Quantitative real-time PCR was performed using real-time thermocycler (Applied Biosystems-7500, USA). RPLP0 was utilized as a housekeeping gene. The primer sequences were 5'-ACTGACATTCTCCACTTCT-3' and 5'-CTTCTGACGCACACCTAT-3' for P53, 5'-GGGACAGCAGCAGGAAGAC-3' and 5'-

GCGTTTGGAGTGGTAGAAA-3' for p21, 5'-GCTCCGTGTTTGGTCAGTGG -3' and 5'-GATGTGGTCAGGGTAGATGGGT-3' for mdm2 and 5'- CCCTGAAGTGCTTGATA -3' and 5'- CCGTTGATGATAGAATGG -3' for RPLP0 genes. The thermal program was a denaturation step for 10 min at 95°C, followed by 40 cycles of 95°C for 10 sec, annealing (59.4 °C for P53,63 °C for p21,53.3 °C for mdm2 and 58.4 °C for RPLP0) for 15 sec, and 72°C for 30 sec.¹¹

P53 transcriptional activity inhibition

To investigate the possible relationship between the observed apoptosis in SW742 cells and the P53 transcriptional pathway, pifithrin (PFT)- α , the inhibitor of P53 transcriptional activity, was used, followed by flow-cytometry analysis. 2.6×105 SW742 cells were seeded into each one of the four 25 cm² (T-25) cell culture flasks and incubated for 12h in the CO2 incubator. Once reaching 70% confluency, the old media were removed and the flasks were treated as follows: one with vehicle (DMSO, 0.1% in media) as the control, one with CSE (1000 µg/ml), and one with Pifithrin- α (30 µM). The fourth flask was treated with Pifithrin- α (30 µM) for 1 h, followed by CSE treatment (1000 µg/ml), and incubated in a 5% CO2 incubator at 37° C for 72h.¹² To evaluate the P53 transcriptional activity inhibition, PI staining/Flow Cytometry was employed. The cells were stained with PI staining solution at 20 µg/ml concentration followed by flow-cytometry according to Riccardi et al.'s method.¹³ Following the incubation time, the fluorescence intensities were read with Becton Dickinson (FACS Calibur, USA) and the results were analyzed via FLOWJO software (version 7.6, USA).

Statistical analysis

All the experiments were carried out in three independent experiments and the data were presented as mean \pm SEM. Statistical analyses were performed using SPSS 17 software (Chicago, USA). We compared the groups with one way ANOVA and LSD, as a post hoc test. *P* values \geq 0.05 were considered to be significant.

Results

The effects of CSE on the expression of protein and mRNA of P53 in SW742 colon cancer cell line

As shown in figure 1, CSE induced the expression of P53 protein in SW742 cells. Western blot analysis of P53 protein expression after 72 h of incubation with CSE showed that this

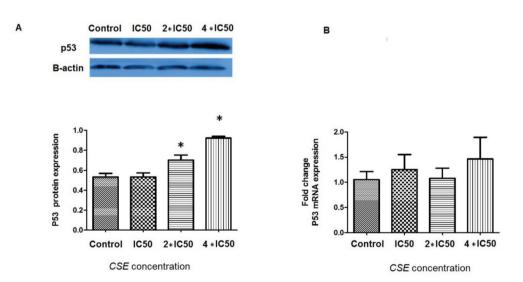


Figure 1. The expression of P53 protein and gene in SW742 treated with CSE. (A) Western blot analysis of P53 protein and its quantification in the control and CSE-treated groups after 72h of treatment. The graph represents the mean values \pm SEM of three independent experiments. (B) Quantitative RT-PCR of P53 gene in the control and CSE-treated groups after 72h of treatment. The graph represents the mean values \pm SEM of three independent experiments.* Significant vs. control (*P*<0.05). CSE: *Cyrtopodion scabrum* extract; RT-PCR: Real time polymerase chain reaction

induction was significant in the $2 \times IC50$ and $4 \times IC50$ of the extract, yet insignificant in IC50 ($251\pm13 \mu g/ml$) compared to the control untreated cells (Figure 1A).

According to the results summarized in figure 1B, P53 mRNA expression level showed no significant changes in different CSE concentrations.

The effects of CSE on the gene expression of two P53 target genes, p21 and mdm2, in SW742 colon cancer cell line

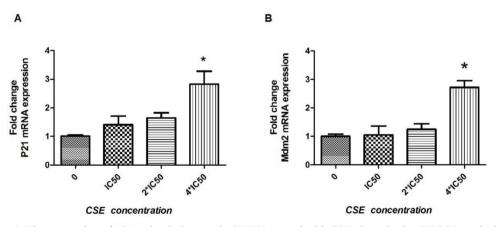
Both p21 and mdm2 mRNA expression level significantly increased at $4\times$ IC50 (1000µg/ml) of CSE compared to the control (~2.8 and 2.7-times, respectively; *P*<0.05) (Figures 2A and 2B). The increase in p21 and mdm2 mRNA expression levels was not significant in the IC50 and 2×IC50 of CSE.

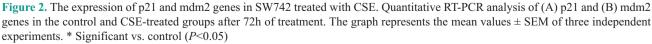
The relationship between apoptosis and P53 transcription pathway

Considering the fact that the Sub-G1 fraction percentage represents apoptotic cells, in this part of the experiment, the percentage of apoptosis was checked with and without PFT- α in SW742 cells treated with CSE. Figure 3 summarizes the results of flow-cytometry analysis. As could be seen, the percentage of Sub-G1 fraction in the control and PFT- α treated groups was the same, showing that this concentration of PFT- α was not toxic to the cells. The comparison of the profile of SW742 cells treated with CSE alone or pretreated with PFT- α revealed that treatment with the combination of PFT- α and the extract had no significant effects on the Sub-G1 fraction percentage. However, treatment with both the extract and/or combination treatment (PFT- α and the extract) demonstrated a significant increase in Sub-G1 fraction percentage compared with the controls. The data suggested that the induction of apoptosis via CSE does not occur through the P53-dependent transcriptional pathway.

Discussion

In the present study, aiming to clarify the mechanism of action of CSE on apoptosis induction and the involvement of the P53 transcriptional pathway in this process, we applied SW742 colorectal cancer cells and performed a series of in vitro experiments. Our findings indicated that the treatment of SW742 cells with different concentrations of CSE leads to increased expression of P53 protein in a dose-dependent manner. While P53 mRNA expression did not show any significant up-regulation, p21 and mdm2 mRNA expression, as two important P53 target genes, were upregulated significantly, particularly at the highest concentration $(1000 \,\mu g/ml)$ of CSE. The upregulation of p21 mRNA was compatible with the results of our previous studies, which implied cell cycle arrest as the result of treatment of SW742 cells with CSE. Furthermore, flowcytometry analysis with PI staining did not show





CSE: Cyrtopodion scabrum extract; RT-PCR: Real time polymerase chain reaction

any significant effects on the subG1 population as the representative of apoptotic cells, following the inhibition of P53 transcription activity with Pifithrin (PFT)- α and CSE.

In the present study, we utilized SW742 cells since it was the most inhibited cell line among digestive cancer cells treated with CSE, according to our previous work. The IC50 value for CSE was also calculated as $251\pm13 \mu g/ml$. According to the data obtained in this part, the expression of TP53 increased in SW742 cells treated with different concentrations (IC50, 2×IC50, and 4×IC50) of CSE in a dose-dependent manner. This upregulation was significant in the cells treated with 2×IC50 (1.5 fold) and 4×IC50 (1.8 fold) of CSE compared with the control untreated cells. These results are in agreement with those obtained from apoptosis assessment in our previous study. The results of P53 mRNA expression revealed that it increased neither accordingly nor significantly in comparison to its protein, TP53. Such inconsistency between P53 protein and mRNA expression has also been reported previously;¹⁵ it has been mentioned that "the induction of the P53 response upon stress occurs largely through alterations in the P53 protein and the changes in the rate of P53 transcription play a minor role, or no roles, in such induction".¹⁶ P53 is known as one of the key tumor suppressor proteins that plays essential roles in appropriate progression of the cell cycle, thereby having the main role in cancer prevention.¹⁴ Herein, we demonstrated that CSE treatment affects the P53 protein pathway in SW742 cells through upregulating the P53 tumor suppressor protein (TP53) expression.

P53 plays its role through the downstream genes, including p21 and mdm2, which results in cell cycle arrest and apoptosis. The results of the expression pattern profiling of p21 gene showed a significant increase in 4×IC50 (1000 μ g /ml) and not at IC50 or 2×IC50 of CSE. As mentioned above, at IC50, the P53 protein level was not significantly upregulated compared to the controls; therefore, no increase in p21 mRNA was expected. However, at 2×IC50, despite the significant increase in the P53 protein level, the p21 mRNA level did not increase significantly. This result is in agreement with that of a study which showed the same inconsistency.¹⁵ However, it seems that a higher level of CSE and thereupon, is needed for induction of p21 mRNA expression, as seen at 4×IC50, with about 2.8 fold upregulation compared with the untreated control cells. This significant induction of p21 gene is in line with previous reports about cell cycle analysis and G2

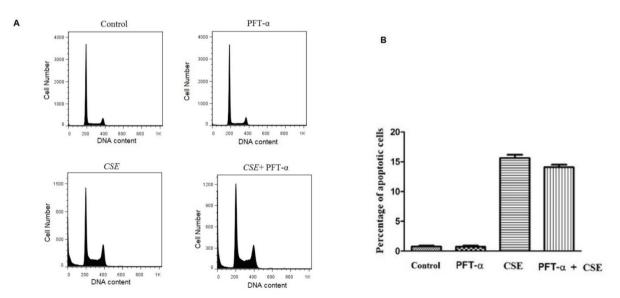


Figure 3. Flow-cytometry analysis of the apoptotic SW742 cells treated with CSE and PFT- α . Controls, PFT- α , and CSE groups were treated with vehicle (DMSO, 0.1% in media), PFT- α (30µM), and CSE (1000 µg/ml), respectively. For PFT- α +CSE, the cells were pre-treated with 30 µM of PFT- α for 1 h, then incubated with CSE for 72 h. The obtained data are presented as mean values ± SEM of three independent experiments.*Significant vs. control (*P*<0.05).CSE: *Cyrtopodion scabrum extract*, PFT- α : pifithrin– β

arrest of SW742 cells at the concentration of 4×IC50.8 p21(Waf1/Cip1) is an important target gene of P53,^{17, 18} which helps regulation of the cell cycle. P53 can transcriptionally upregulate p21,19 which inhibits G1/S and G2/M phases; thus, P53 is capable of controlling both G1 and G2/M checkpoints through p21 function. Interestingly, the same profile was observed for mdm2 gene expression; accordingly, at the concentration of 4×IC50, about 2.7 fold upregulation was observed compared with the control cells. In unstressed conditions, mdm2, as a P53 target gene, provided a negative feedback loop; therefore, its gene product, MDM2 protein, binds to and efficiently inhibits P53 protein.²¹ Mdm2 plays this role in two major ways: binding to the transactivation domain of P5322 and acting as an E3 ubiquitin ligase,²³ leading to changes in the localization and proteasomal degradation of TP53. In stressed conditions, the interaction between P53 and MDM2 will be disrupted through certain mechanisms, including P53 and MDM2 phosphorylation and sequestration of MDM2 by p14 ARF.²⁴ The results of this section revealed that the CSE at the concentration of 1000 µg/ml (4×IC50) plays a positive role in the inhibition of cell proliferation by upregulating these two genes as the consequence of P53 upregulation.

The last part of this study was designed to investigate whether apoptosis occurs through P53 transcription pathway. The findings implied that blocking P53 transcriptional activity with PFT- α in SW742 cells treated with CSE did not affect the percentage of apoptotic cells. Pifithrin (PFT)- α is a small molecule that binds to the DNA binding domain of P53. It also blocks the interaction of P53 with the transcriptional cofactor p300, thereby inhibiting P53 transcriptional activity.¹² The name of Pifithrin is rooted in pfifty three inhibitor.²⁵ P53 induces apoptosis through two mechanisms, including transcriptional- dependent and independent mechanisms, such as a direct localization of P53 to the mitochondria.²⁶ Our results are in accordance with those of another study that reported apoptosis induction through the P53 transcriptional independent pathway despite P53 protein elevation.¹² Therefore, CSE-induced apoptosis in SW742 cells might occurs independently of P53 transcriptional activity rather than via a P53 transcription-dependent mechanism.

Conclusion

In conclusion, the present in vitro study partially unfolded the role of CSE in P53 signaling pathway and showed that it induces P53 protein as well as p21 and MDM2 gene expression, which all play pivotal roles in cell cycle arrest. Furthermore, it clarified that induced apoptosis does not occur through the P53 transcriptional pathway. These findings are valuable since we tried to partially explore the mechanism of action of a putative natural anticancer drug which harbors no undesirable effects on normal cells. However, numerous proteins and genes are involved in cancer progression in the cells, which may be affected by CSE treatment, yet were not included in our study. Therefore, further investigation concerning the mechanism of growth suppression is needed. In addition, we could suggest further evaluation of the efficacy of the extract in the animal model, which is fortunately under way in the parallel projects.

The current study is part of a mega project with the ultimate objective of applying CSE in clinical trial and the evaluation of its efficacy and safety as complementary and/or alternative medicine for patients suffering from cancer.

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Conflict of Interest

None declared.

References

- Sudhakar A. History of cancer, ancient and modern treatment methods. *J Cancer Sci Ther.* 2009;1(2):1-4. doi:10.4172/1948-5956.100000e2.
- 2. Hakim L, Alias E, Makpol S, Ngah WZ, Morad NA, Yusof YA. Gelam honey and ginger potentiate the anti

cancer effect of 5-FU against HCT 116 colorectal cancer cells. *Asian Pac J Cancer Prev*. 2014;15(11):4651-7. doi:10.7314/apjcp.2014.15.11. 4651.

- Akhavan A, Binesh F, Soltani A. Survival of rectal cancer in Yazd, Iran. Asian Pac J Cancer Prev. 2014;15(12):4857-60. doi:10.7314/apjcp.2014.15. 12.4857.
- Jeong AJ, Chung CN, Kim HJ, Bae KS, Choi S, Jun WJ, et al. Gecko proteins exert anti-tumor effect against cervical cancer cells via PI3-Kinase/Akt pathway. *Korean J Physiol Pharmacol.* 2012;16(5):361-5. doi:10.4196/kjpp.2012.16.5.361.
- Lee JE, Lee DG, Park SY, Jo A, Kim HK, Han J, et al. Gekkonidae, Lizard tail extracts elicit apoptotic response against non-small cell lung cancer via inhibiting Akt signaling. *Biomed Pharmacother*. 2019;116:109050. doi: 10.1016/j.biopha.2019.109050.
- Duan YM, Jin Y, Guo ML, Duan LX, Wang JG. Differentially expressed genes of HepG2 cells treated with gecko polypeptide mixture. *J Cancer*. 2018;9(15):2723. doi: 10.7150/jca.26339.
- Amiri A, Namavari M, Rashidi M, Fahmidehkar MA, Seghatoleslam A. Inhibitory effects of *Cyrtopodion* scabrum extract on growth of human breast and colorectal cancer cells. Asian Pac J Cancer Prev. 2015;16(2):565-70. doi: 10.7314/apjcp.2015.16.2.565.
- Rashidi M, Seghatoleslam A, Namavari M, Amiri A, Fahmidehkar MA, Ramezani A, et al. Selective Cytotoxicity and apoptosis-induction of *Cyrtopodion scabrum* extract against digestive cancer cell lines. *Int J Cancer Mana*. 2017. doi: 10.5812/ijcm.8633.
- 9. Rastegar-Pouyani N, Khosravani A, Oraie H. A new record of *Cyrtopodion scabrum* (Heyden, 1827) from the Caspian Sea Coastal Region, Guilan Province, Northern Iran. *Herpetology Notes*. 2010;3:61-3.
- Sajadi MM, Mansouri D, Sajadi M-RM. Ibn Sina and the clinical trial. *Ann Intern Med.* 2009;150(9):640-3. doi: 10.7326/0003-4819-150-9-200905050-00011.
- 11. Schmittgen TD, Livak KJ. Analyzing real-time PCR data by the comparative CT method. *Nat Protoc*. 2008;3(6):1101-8. doi: 10.1038/nprot.2008.73.
- Wong FC, Woo CC, Hsu A, Tan BKH. The anti-cancer activities of Vernonia amygdalina extract in human breast cancer cell lines are mediated through caspasedependent and p53-independent pathways. *PLoS One.* 2013;8(10):e78021. doi: 10.1371/journal.pone. 0078021.
- Riccardi C, Nicoletti I. Analysis of apoptosis by propidium iodide staining and flow cytometry. *Nat Protoc.* 2006;1(3):1458-61. doi: 10.1038/nprot.2006. 238.
- 14. Levine AJ, Oren M. The first 30 years of p53: growing ever more complex. *Nat Rev Cancer*. 2009;9(10):749-58. doi: 10.1038/nrc2723.
- 15. McVean M, Xiao H, Isobe KI, Pelling JC. Increase in

wild-type p53 stability and transactivational activity by the chemopreventive agent apigenin in keratinocytes. *Carcinogenesis*. 2000;21(4):633-9. doi: 10.1093/carcin/21.4.633.

- Oren M. Regulation of the p53 tumor suppressor protein. *J Biol Chem.* 1999;274(51):36031-4. doi: 10.1074/jbc.274.51.36031.
- El-Deiry WS, Tokino T, Velculescu VE, Levy DB, Parsons R, Trent JM, et al. WAF1, a potential mediator of p53 tumor suppression. *Cell*. 1993;75(4):817-25. doi: 10.1016/0092-8674(93)90500-p.
- Harper JW, Adami GR, Wei N, Keyomarsi K, Elledge SJ. The p21 Cdk-interacting protein Cip1 is a potent inhibitor of G1 cyclin-dependent kinases. *Cell*. 1993;75(4):805-16. doi: 10.1016/0092-8674(93)90499-g.
- 19. Chen J, Pan J, Wang J, Song K, Zhu D, Huang C, et al. Soluble egg antigens of Schistosoma japonicum induce senescence in activated hepatic stellate cells by activation of the STAT3/p53/p21 pathway. *Sci Rep.* 2016; 6:30957. doi: 10.1038/srep30957.
- 20. Agarwal ML, Agarwal A, Taylor WR, Stark GR. p53 controls both the G2/M and the G1 cell cycle checkpoints and mediates reversible growth arrest in human fibroblasts. *Proc Natl Acad Sci.* 1995;92(18):8493-7. doi: 10.1073/pnas.92.18.8493.
- 21. Momand J, Zambetti GP, Olson DC, George D, Levine AJ. The mdm-2 oncogene product forms a complex with the p53 protein and inhibits p53-mediated transactivation. *Cell*. 1992; 69(7):1237-45. doi: 10.1016/0092-8674(92)90644-r.
- Oliner JD, Pietenpol JA, Thiagalingam S, Gyuris J, Kinzler KW, Vogelstein B. Oncoprotein MDM2 conceals the activation domain of tumour suppressor p53. *Nature*. 1993: 362(6423):857-60. doi: 10.1038/362857a0.
- 23. Kubbutat MH, Jones SN, Vousden KH. Regulation of p53 stability by Mdm2. *Nature*. 1997;387(6630):299-303. doi: 10.1038/387299a0.
- 24. Scoumanne A, Harms KL, Chen X. Structural basis for gene activation by p53 family members. *Cancer Biol Ther.* 2005;4(11):1178-85. doi: 10.4161/cbt.4.11. 2254.
- Charlot J, Nicolier M, Pretet J, Mougin C. Modulation of p53 transcriptional activity by PRIMA-1 and Pifithrin-? on staurosporine-induced apoptosis of wildtype and mutated p53 epithelial cells. *Apoptosis*. 2006;11(5):813-27. doi: 10.1007/s10495-006-5876-6.
- Moll UM, Wolff S, Speidel D, Deppert W. Transcription-independent pro-apoptotic functions of p53. *Curr Opin Cell Biol.* 2005;17(6):631-6. doi: 10.1016/j.ceb.2005.09.007.